Inhibition of MEK/ERK/STAT3 signaling in oleuropein treatment inhibits myocardial ischemia/reperfusion

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Abstract. Studies have shown that oleuropein has antifungal, anti-inflammatory, antiviral, antioxidant, anticancer and hypoglycemic functions. TTC solution staining was used to measure myocardial infarction size. A commercial kit was used to measure lactate dehydrogenase (LDH), creatinine kinase-MB (CK-MB), tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), IL 6, superoxide dismutase (SOD), glutathione (GSH), malondialdehyde (MDA) and catalase levels. Western blot analysis was used to measure p53, p-MEK p-ERK and p-IκBα protein expression. The present study reports that the protective effect of oleuropein also prevents against myocardial ischemia/reperfusion (myocardial I/R). The aim of this retrospective study was to evaluate this protective effect of oleuropein and the mechanisms by which myocardial I/R is prevented. Oleuropein inhibited myocardial infarction size, CK-MB and LDH serum levels in a myocardial I/R rat model. Moreover, oleuropein also attenuated caspase-3 activity, and p53, phosphorylated (p)-mitogen-activated protein kinase (MEK), p-extracellular signal-regulated protein kinase (ERK) and p-IκBα protein expression. TNF-α, IL-1β, IL-6 and MDA were decreased; SOD, GSH and catalase levels inhibited TNF-α, IL-1β, IL-6 and MDA levels, and increased SOD, GSH and catalase levels in myocardial I/R rats treated with oleuropein. Rats orally administered the MEK inhibitor PD0325901, in addition to oleuropein, exhibited inhibited myocardial infarction size, CK-MB and LDH serum levels compared with rats treated with oleuropein only. Rats treated with MEK inhibitor also exhibited suppressed caspase-3 activity, p53, p-MEK p-ERK and p-IκBα protein expression, TNF-α, IL-1β, IL-6, SOD, GSH, MDA and catalase levels, and induced p-signal transducer and activator of transcription 3 (STAT3) protein expression compared with rats treated with oleuropein only. Taken together, these results suggest that MEK/ERK/STAT3 signaling regulates the inhibition of myocardial I/R in rats treated with oleuropein.

Introduction

With the continuous improvement of living standards and the accelerated pace of life, the morbidity and mortality rates of coronary atherosclerotic heart disease are increasing (1). The World Health Organization has indicated that coronary heart disease has become the leading cause of mortality in the world, and that acute myocardial infarction (AMI) is the leading cause of coronary heart disease-related mortality (2). With the continuous development of treatment, the morbidity rate of heart failure resulting from ventricular remodeling and of arrhythmia due to sympathetic remodeling is gradually increasing, while the acute-phase mortality rate of AMI is gradually decreasing (3,4). Even though patients may receive optimized medical treatment, the overall mortality rate remains high.

Myocardial apoptosis is an autonomous process of programmed cell death in a series of gene regulation, of which the essence is physiological cell death (5). Myocardial apoptosis exists in the cardiovascular system, particularly during the physiological and pathological changes of AMI, and is an important cellular basis for a variety of cardiovascular diseases (6). The inhibition or reduction of myocardial apoptosis can reflect ventricular remodeling and cardiac function recovery following AMI, so the study of myocardial apoptosis is essential to evaluate the post-myocardial infarction heart function and associated drug efficacy (7).

AMI leads to an abnormal cellular environment due to an insufficient energy supply. The increase in cardiac compensatory contraction results in an elevated reactive oxygen species (ROS) level caused by membrane nicotinamide adenine dinucleotide phosphate (8). More seriously, the elevated ROS level triggers mitochondria to generate a large amount of oxidative stress (9). Oxidative stress can not only attack the cell membrane and organelles, but also cause the...
inflammatory response through mutual reinforcement with inflammatory cytokines, to further aggravate myocardial injury caused by myocardial infarction (10).

Heart inflammation can be summarized as a simple natural immune response and/or the combination of a natural immune response and an acquired immune response (11). The most typical characteristic of a natural immune response is the induction of inflammatory cytokine generation. In myocardial ischemia and heart failure, a natural immune response and an inflammatory response usually occur (12).

Oleuropein (Fig. 1) is a non-toxic split iridoid glycoside compound. Oleuropein increases the coronary blood flow of the rabbit isolated heart by 50%, indicating antiarrhythmic and anti-spasmodic effects (13). Oleuropein obtained by the hydrolysis of olive leaf extract has antihypertensive effect (14). In addition, oleuropein is a strong angiotensin-converting enzyme inhibitor, for which the inhibitory effect is a result of the inherent 2,3-dihydroxy isoleucine (15). The corresponding aglycone produced by the enzymatic hydrolysis shows a similar effect to oleuropein, which has long-lasting hypotensive effect on rats, cats and dogs (16). Therefore, the present study aimed to investigate the protective effect and mechanisms of oleuropein in myocardial ischemia/reperfusion (I/R), and the possible role of extracellular signal-regulated protein kinase (ERK) signaling in the protective effects of oleuropein in myocardial I/R injury.

Materials and methods

Animals. All procedures were performed in accordance with the National Research Council's Guide of HARRISON International Peace Hospital, Hebei Medical University (Hengshu, Hebei, China) for Humane Care and Use of Laboratory Animals. All animal experiments were approved by the Medical Ethics Committee of HARRISON International Peace Hospital. Adult, male, Sprague-Dawley rats weighing 250-300 g were purchased from Hebei Medical University and maintained in controlled conditions of 22±2˚C and 60-70% humidity under a 12 h light-dark cycle (7:00 a.m. to 7:00 p.m.). Food and water were available ad libitum.

In vivo myocardial I/R model and experimental groups. Firstly, a total of 26 rats (age, 5-6 weeks; weight, 200-250 g) were randomized into 3 groups: i) Control group (n=6): rats subjected to the surgical procedures without coronary occlusion; ii) myocardial I/R model group (n=10): 30 min of coronary occlusion followed by 3 h of reperfusion; and iii) Ole (20) group (n=10): 20 mg/kg of oleuropein for 2 consecutive days, then 30 min of coronary occlusion followed by 3 h of reperfusion.

Next, a total of 36 rats (age, 5-6 weeks; weight, 200-250 g) were randomized into 4 groups: i) Control group (n=6): rats subjected to the surgical procedures without coronary occlusion; ii) myocardial I/R model group (n=10): 30 min of coronary occlusion followed by 3 h of reperfusion; iii) Ole (20) group (n=10): 20 mg/kg of oleuropein for 2 consecutive days, then 30 min of coronary occlusion followed by 3 h of reperfusion; and iv) PD0325901 group: 3 mg/kg of PD0325901 and 20 mg/kg of oleuropein for 2 consecutive days, then 30 min of coronary occlusion followed by 3 h of reperfusion.

Rats were anesthetized by an intraperitoneal injection of 100 mg/kg ketamine (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). A left thoracotomy was performed to expose the hearts and the left anterior descending (LAD) artery was ligated at 2-3 mm at the pulmonary artery conus. Next, the left atrium was sutured using 6-0 silk Prolene. For reversible coronary occlusion, a small vinyl tube was placed on top of the vessel to form a snare. Regional ischemia was sustained in the heart for 30 min and reperfusion was then performed with release of the slipknot. Rats were sacrificed using decollation under anesthesia and the heart was peeled and frozen at 80˚C.

Measurement of myocardial infarction size. Following reperfusion for 3 h, the LAD artery was removed and then 2% Evans blue (1.8-2 ml) was injected intravenously to denote the area at risk. The heart was peeled and frozen at -80˚C for 24 h. The heart was then vertically cut into 1.5-mm sections from the long axis to the area of ligation. Sections were incubated in 1% TTC solution for 30 min at 37˚C and then incubated with 10% neutral buffered formalin overnight at room temperature.

Measurements of lactate dehydrogenase (LDH), creatinine kinase-MB (CK-MB), tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), IL-6, superoxide dismutase (SOD), glutathione (GSH), malondialdehyde (MDA) and catalase levels. Subsequent to reperfusion, blood samples were collected from the right ventricle of every rat. These were centrifuged at 3,000 x g for 10 min at 4˚C to separate the serum. LDH and CK-MB levels were evaluated using commercially available assay kits (Sigma-Aldrich; Merck KGaA), TNF-α, IL-1β, IL-6, SOD, GSH, MDA and catalase levels were evaluated using commercial enzyme-linked immunosororbent assay (ELISA) assay kits (Wuhan Boster Biological Technology, Inc., Wuhan, China).

Determination of apoptosis. Cytoplasmic proteins were prepared from heart tissues following reperfusion for 3 h and were lysed in ice-cold extraction buffer containing protease inhibitor cocktail (both Beyotime Institute of Technology, Shanghai, China) for 30 min. Miscible liquids were centrifuged at 12,000 x g for 30 min and levels determined using a modified Bradford assay (Bio-Rad Laboratories, Inc., Hercules, CA, USA). Protein (10 µg) was used to measure the activity levels of caspase-3 using caspase-3 activity kit (Beyotime Institute of Technology).
Western blot analysis. Cytoplasmic proteins were prepared from heart tissues following reperfusion for 3 h and were lysed in ice-cold extraction buffer containing protease inhibitor cocktail (both Beyotime Institute of Technology) for 30 min. Miscible liquids were centrifuged at 12,000 x g for 30 min and levels determined using a modified Bradford assay (Bio-Rad Laboratories, Inc.). Protein (50-60 µg) was separated by electrophoresis on 10% sodium dodecyl sulfate-polyacrylamide gels and transferred to nitrocellulose membranes (Bio-Rad Laboratories, Inc.). Membranes were washed with 5% bovine serum albumin with Tris-buffered saline (TBS; 0.01 M, pH 7.4) for 1 h at 37˚C and then incubated with peroxidase-conjugated secondary antibodies (sc-2004; 1:5,000 dilution, Santa Cruz Biotechnology, Inc.) for 1 h at 37˚C. The signals were detected with the enhanced chemiluminescence system (GE Healthcare, Chicago, IL, USA) and assayed by Image_Lab_3.0 (Bio-Rad Laboratories, Inc.).

Statistical analysis. Data are presented as the mean ± standard error. Data were analyzed using StatSoft Statistica version 13.0 (StatSoft Inc., Tulsa, OK, USA). Statistical analyses were performed using one-way analysis of variance with repeated measures, followed by Bonferroni’s post-hoc test. Statistical significance was defined as P<0.05.

Results

Protective effect of oleuropein against myocardial infarction size in the myocardial I/R rats. Compared with rats in the normal control group, rats in the myocardial I/R group exhibited a significant increase in myocardial infarction size (Fig. 2). Administration of oleuropein significantly inhibited the induction of myocardial infarction size by myocardial I/R injury compared with the myocardial I/R model (Fig. 2).

Protective effect of oleuropein against CK-MB and LDH serum levels in myocardial I/R rats. CK-MB and LDH levels in the serum were also examined as indicators for myocardial injury evaluation in the present study. Compared with levels in the normal control group, there was a significant increase in CK-MB and LDH serum levels in the myocardial I/R rat group (Fig. 3). Consistently, rats with oleuropein treatment exhibited reduced levels of CK-MB and LDH compared with rats in the myocardial I/R group (Fig. 3).

Protective effect of oleuropein against caspase-3 activity expression in myocardial I/R rats. To research the protective effect of oleuropein against apoptosis, caspase-3 activity expression in the myocardial I/R rat was researched. In comparison with that in the normal control group, a significant induction in caspase-3 activity was found in the myocardial I/R rat group (Fig. 4). However, rats with oleuropein administration exhibited significantly inhibited caspase-3 activity compared with rats in the myocardial I/R group (Fig. 4).

Protective effect of oleuropein against p53, MEK and ERK protein expression in myocardial I/R rats. To further research
the mechanism behind the protective effect of oleuropein against apoptosis, p53, MEK and ERK protein expression in myocardial I/R rats was measured using western blot analysis. The induction of p53, p-MEK and p-ERK protein expression was markedly observed in the myocardial I/R model group in comparison with that in the normal control group (Fig. 5). The oleuropein treatment group showed significantly suppressed induction of p53, p-MEK and p-ERK protein expression compared with the myocardial I/R group.

Protective effect of oleuropein against p-IκBα protein expression in myocardial I/R rats. To investigate the mechanism behind the protective effect of oleuropein against inflammation factors, p-IκBα protein expression in myocardial I/R rats was measured using western blot analysis. Myocardial I/R significantly induced the p-IκBα protein expression in the myocardial I/R rat group compared with that in the normal control group (Fig. 6). The oleuropein-treated group showed significant suppression of p-IκBα protein expression compared with the myocardial I/R rat group (Fig. 6).

Protective effect of oleuropein against TNF-α, IL-1β and IL-6 levels in myocardial I/R rats. To investigate the anti-inflammatory effect of oleuropein against myocardial I/R, TNF-α, IL-1β and IL-6 levels were measured using ELISA assay kits. The results demonstrated that the TNF-α, IL-1β and IL-6 levels were significantly induced in the myocardial I/R model

Figure 5. Protective effect of oleuropein against p53, MEK and ERK protein expression in myocardial I/R rats, as determined using (A) western blotting and statistical analysis of (B) p53, (C) p-MEK and (D) p-ERK protein expression. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day oleuropein treatment group. **P<0.01 vs. control group; *P<0.01 vs. myocardial I/R group. Ole, oleuropein; I/R, ischemia/reperfusion; ER, extracellular signal-regulated protein kinase; MEK, mitogen-activated protein kinase kinase; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; p-, phosphorylated.

Figure 6. Protective effect of oleuropein against p-IκBα protein expression, as determined using (A) western blotting and (B) statistical analysis of p-IκBα protein expression in myocardial I/R rats. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day oleuropein treatment group. **P<0.01 vs. control group; *P<0.01 vs. myocardial I/R group. Ole, oleuropein; I/R, ischemia/reperfusion; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; p-, phosphorylated.

Figure 7. Protective effect of oleuropein against (A) TNF-α, (B) IL-1β and (C) IL-6 levels in myocardial I/R rats. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day oleuropein treatment group. **P<0.01 vs. control group; *P<0.01 vs. myocardial I/R group. Ole, oleuropein; I/R, ischemia/reperfusion; IL, interleukin; TNF-α, tumor necrosis factor-α.
The oleuropein-treated group exhibited significantly reduced induction of TNF-α, IL-1β and IL-6 levels by myocardial I/R compared with the myocardial I/R group (Fig. 7). The oleuropein-treated group exhibited significantly increased levels of SOD and GSH, and a significantly decreased MDA level compared with rats in the myocardial I/R model group (Fig. 8).

Protective effect of oleuropein on p-STAT3 protein expression in myocardial I/R rats. Additionally, the protective effect of oleuropein against p-STAT3 (p-STAT3) protein expression was investigated in the myocardial I/R rat. As shown in Fig. 9, the suppression of p-STAT3 protein expression was markedly increased in the myocardial I/R rat group compared with that in the normal control group. Rats treated with oleuropein exhibited significantly induced p-STAT3 protein expression compared with rats in the myocardial I/R group (Fig. 9).

Inhibition of MEK increases the protective effect of oleuropein against myocardial infarction size in myocardial I/R rats. To provide evidence linking MEK to the effect of oleuropein, PD0325901, a MEK inhibitor, was introduced to the myocardial I/R rats to analyze how the inhibition of MEK affected
the protective effect of oleuropein against myocardial infarction size. The group treated with PD0325901 and oleuropein exhibited a significantly inhibited myocardial infarction size compared with the group treated with oleuropein only (Fig. 10).

**Inhibition of MEK increases the protective effect of oleuropein against CK-MB and LDH serum levels in myocardial I/R rats.** To investigate the mechanism of oleuropein against myocardial I/R, CK-MB and LDH serum levels were examined in myocardial I/R rats treated with oleuropein following inhibition of MEK. Rats with MEK inhibition exhibited significantly decreased CK-MB and LDH serum levels compared with rats treated with oleuropein only (Fig. 11).

**Inhibition of MEK increases the protective effect of oleuropein against caspase-3 activity expression in myocardial I/R rats.** To determine whether the inhibition of MEK affects apoptosis in oleuropein-treated myocardial I/R rats, caspase-3 activity expression was examined after the addition of PD0325901. Rats with MEK inhibition showed significantly decreased caspase-3 activity compared with rats treated with oleuropein only (Fig. 12).

**Inhibition of MEK increases the protective effect of oleuropein against p53, MEK and ERK protein expression in myocardial I/R rats.** To further determine whether the inhibition of MEK regulates p53, MEK and ERK protein expression in myocardial I/R rats treated with oleuropein, protein levels were analyzed. Following MEK inhibition, the protein expression of p53, p-MEK and p-ERK was significantly suppressed compared with that in the group treated with oleuropein only (Fig. 13A-D).

**Inhibition of MEK increases the protective effect of oleuropein against p-IkBα protein expression in myocardial I/R rats.** To determine whether the inhibition of MEK increased the anti-inflammatory effect of oleuropein in myocardial I/R rats, p-IkBα protein expression was detected using western blot analysis. Following MEK inhibition, the protein expression of p-IkBα was also significantly suppressed compared with that in the group treated with oleuropein only (Fig. 13A and E).

**Inhibition of MEK increases the protective effect of oleuropein against TNF-α, IL-1β and IL-6 levels in myocardial I/R rats.** To further determine whether the inhibition of MEK affects the anti-inflammatory effect of oleuropein in myocardial I/R rats, TNF-α, IL-1β and IL-6 levels were analyzed. It was found that the inhibition of MEK significantly inhibited TNF-α, IL-1β and IL-6 levels in the rats treated with PD0325901 and oleuropein compared with that in rats treated with oleuropein only (Fig. 14).

**Inhibition of MEK increases the protective effect of oleuropein against SOD, GSH and MDA levels in myocardial I/R rats.** Next, the effect of the inhibition of MEK on the anti-oxidative action of oleuropein was examined in the myocardial I/R rats. With the inhibition of MEK, a significant increase in SOD and GSH levels, and a significant decrease in MDA level was found in the PD0325901 group compared with that in the group treated with oleuropein only (Fig. 15).

**Inhibition of MEK increases the protective effect of oleuropein on p-STAT3 in myocardial I/R rats.** Lastly, whether the inhibition of MEK increased the protective effect of oleuropein against p-STAT3 was examined. The protein expression of p-STAT3 was significantly promoted in the PD0325901 group compared with that in the group treated with oleuropein only (Fig. 16).

**Discussion**

Myocardial ischemia refers to the reduction of heart blood perfusion, resulting in cardiac oxygen reduction and abnormal myocardial energy metabolism, which is a pathological state that cannot support heart function (1). Coronary plaques or occlusion and instability of coronary atherosclerosis caused by coronary artery stenosis are major causes of myocardial ischemia (17). Patients with recurrent myocardial ischemia may suffer from angina, myocardial stunning, myocardial hibernation, ischemic preconditioning, acute coronary syndrome, AMI or even cardiac rupture (18). In the present study, oleuropein significantly inhibited the myocardial infarction size and reduced the levels of CK-MB and LDH in the myocardial I/R rat. Nekooeian et al (15) reported that oleuropein offered cardioprotection via its antioxidant properties.

ERK1/2 is a subfamily of the mitogen-activated protein kinase (MAPK) signaling pathway, which exists widely in various stages of the cell cycle and serves an important role in gene transcription and cell cycle processes (19). At present, ERK1/2 is regarded to promote both cell survival and proliferation, and apoptosis, for which the mechanism depends on the sub-localization of ERK1/2 in cells and the activated signaling molecules downstream (19). According to the traditional view, ERK1/2 enters into the nucleus to activate cell survival and proliferation; however, if the phosphorylated ERK1/2 stays in the cytoplasm for the long term, it may interact with a series of pro-apoptotic proteins to initiate the apoptosis of cells (20). The present study found that oleuropein significantly suppressed the induction of p53, p-MEK and p-ERK protein expression in myocardial I/R rats. Potočnjak et al (21) suggest that oleuropein attenuated cisplatin-induced acute renal injury through inhibition of p53 and ERK signaling in mice.

Cardiac cells are a class of mature cells with terminal differentiation and without the proliferation ability (22).
Figure 13. Inhibition of MEK increases the protective effect of oleuropein against p53, MEK, ERK and p-IκBα protein expression, as determined using western blotting (A) and statistical analysis of (B) p53, (C) p-MEK, (D) p-ERK and (E) p-IκBα protein expression in myocardial I/R rats. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day of oleuropein treated group; PD0325901, 3 mg/kg of PD0325901 + 20 mg/kg/day of oleuropein treated group. **p<0.01 vs. the control group; ***p<0.01 vs. the myocardial I/R group; ****p<0.01 vs. Ole (20) group. Ole, oleuropein; I/R, ischemia/reperfusion; p-, phosphorylated; MEK, mitogen-activated protein kinase kinase; ERK, extracellular signal-regulated protein kinase; GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

Figure 14. Inhibition of mitogen-activated protein kinase kinase increases the protective effect of oleuropein against (A) TNF-α, (B) IL-1β and (C) IL-6 levels in myocardial I/R rats. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day oleuropein treatment group; PD0325901, 3 mg/kg PD0325901 + 20 mg/kg/day oleuropein treatment group. **P<0.01 vs. control group; ***P<0.01 vs. myocardial I/R group; ****P<0.01 vs. Ole (20) group. Ole, oleuropein; I/R, ischemia/reperfusion; IL, interleukin; TNF-α, tumor necrosis factor-α.

Figure 15. Inhibition of mitogen-activated protein kinase kinase increases the protective effect of oleuropein against (A) SOD, (B) GSH and (C) MDA levels in myocardial I/R rats. Control, control group; myocardial I/R, myocardial I/R model group; Ole (20), 20 mg/kg/day oleuropein treatment group; PD0325901, 3 mg/kg PD0325901 + 20 mg/kg/day oleuropein treatment group. **P<0.01 vs. control group; ***P<0.01 vs. myocardial I/R group; ****P<0.01 vs. Ole (20) group. Ole, oleuropein; I/R, ischemia/reperfusion; SOD, superoxide dismutase; GSH, glutathione; MDA, malondialdehyde; pro, protein.
Myocardial cell apoptosis is a type of programmed cell death that is performed in a series of gene regulation by activating cell ‘suicide’ program through certain signaling pathways under certain physiological and pathological conditions (22). It has been indicated that the apoptosis in myocardial cells serves an important role in the physiological and pathological developmental processes and maintenance of normal heart morphology, and is considered to be the ‘cellular basis’ for the change from compensatory changes to pathological changes (23). Meanwhile, the present study found that oleuropein administration significantly inhibited caspase-3 activity in the myocardial I/R rat. Impellizzeri et al (13) showed that oleuropein may be useful in the treatment of various inflammatory diseases via suppression of caspase-3 in mice with spinal cord injury.

In AMI, long-term hypoxia and ischemia of myocardial cells leads to aerobic metabolic disorder due to coronary occlusion, and then causes hyperemia and edema of myocardial interstitial cells, and myocardial cell degeneration and necrosis, accompanied by a large amount of inflammatory cell infiltration (10). A large number of free radicals are generated in the tissues and the peroxide destruction of oxygen free radicals mainly damages the structure and function of myocardial cell membranes, damages the mitochondria, cuts off the cells energy supply and destroys lysosomes to cause cell autolysis (24). The myocardium of accelerated ischemia develops from reversible damage to irreversible degeneration and necrosis. Malignant arrhythmia appears, thereby causing ventricular remodeling and cardiac dysfunction (24). Recanalization and reperfusion therapy is the most effective treatment now, but I/R can further damage the myocardium, for which the important mechanism is oxidative stress; the greater the duration of myocardial ischemia and hypoxia the greater the oxidative stress and myocardial damage, and the more severe the disease (25). The present study showed that oleuropein significantly increased the inhibition of SOD and GSH, and inhibited the activation of MDA level in myocardial I/R rats. Nekooeian et al (15) indicated that oleuropein offered cardioprotection through its antioxidant properties in rats with simultaneous type 2 diabetes.

AMI is the myocardial necrosis caused by acute and persistent myocardial ischemia and hypoxia. Following AMI, changes such as myocardial ischemia, hypoxia and increases in wall tension among, other others, may induce the formation of the inflammatory response (26). In the inflammatory process subsequent to AMI, IL-6 and TNF-α serve greater roles. IL-6 promotes the increased expression of intercellular adhesion molecule-1 by myocardial cells via the regulation of
the synthesis of liver C-reactive protein (CRP) and a series of biochemical processes of the liver in the acute phase reaction, thereby enhancing the adhesion of neutrophils and the release of oxygen free radicals (27). TNF-α directly damages endothelial cells to induce the CRP synthesis pathway to produce an effect by enhancing leukocyte chemotaxis (27). The aforementioned inflammatory processes will inevitably result in myocardial cell injury or myocardial fibrosis. Thus far, studies have shown that inflammation serves an important role in the development of myocardial fibrosis (28). In the present study, it was found that oleuropein significantly reduced the induction of TNF-α, IL-1β and IL-6 levels by myocardial I/R. Giner et al (16) also showed oleuropein to be a protective agent against colitis-associated colorectal cancer via reduction of intestinal IL-6, IFN-γ, TNF-α and IL-17A concentration in c57bl/6 mice.

In the resting condition of cells, nuclear factor-κB (NF-κB) is bound to IκB to form a complex present in the cytoplasm. When subjected to external stimuli, including cytokines, oxidants, protein kinase C activator, viruses, ultraviolet and lipopolysaccharides, IκBs are degraded, thus releasing the free NF-κB dimers. At this time, the NF-κB is transported from the cytoplasm to the nucleus, which influences the transcription of various adhesion cytokines, immune receptors, acute-phase proteins and stress-response protein genes (29). Studies have shown that NF-κB is a central regulator for stress and the inflammatory response, and that it not only serves a role in immune regulation, but that its signaling pathway has also been extensively involved in cell survival, differentiation, proliferation and apoptosis, playing an important role in the occurrence, development and outcome of numerous diseases (30,31). The present study showed that p-IκB protein expression was significantly suppressed in myocardial I/R rats with oleuropein treatment compared with that in rats without treatment. Campolo et al (32) demonstrated that oleuropein inhibits secondary events of intestinal ischemia/reperfusion injury through NF-κB and IκBα.

ERK is the most important and classical route that is best studied for the MAPK path, the path is the nodes and common pathway of multiple pro-proliferation signal transduction pathways (33). Previous results have shown that angiotensin II (AngII) may lead to pathological myocardial hypertrophy and myocardial fibrosis by AngII type I receptor (AT,R)-mediated protein kinase C (PKC)-ERK1/ERK2 pathway signaling (34). The binding of AngII and AT,R can activate a number of signaling molecules downstream, including phospholipase C, phospholipase D and PKC. PKC activates ERK1/2, thus achieving the signal transduction within the cell and the activation of nuclear gene transcription (35). Notably, in the present study, inhibition of MEK expression could inhibit myocardial infarction size and CK-MB and LDH serum levels, suppress caspase-3 activity and p53, p-MEK p-ERK and p-IκBα protein expression, and inhibit TNF-α, IL-1β, IL-6 and MDA levels and increase SOD, GSH and catalase levels in myocardial I/R rats treated with oleuropein.

In conclusion, the results of the current study showed that oleuropein inhibited myocardial infarction size, and CK-MB and LDH serum levels in myocardial I/R rats through anti-inflammation, anti-oxidant, anti-apoptosis and inhibition of MEK/ERK/STAT3 signaling. Moreover, inhibition of the MEK/ERK/STAT3 signaling pathway may serve a key role in the protective effects of oleuropein against myocardial I/R in rats (Fig. 17).

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Availability of data and materials

The analyzed data sets generated during the study are available from the corresponding author on reasonable request.

Authors’ contributions

HXJ designed the experiment. HXJ analyzed the data and wrote the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

All animal experiments were approved by the Medical Ethics Committee of Harrison International Peace Hospital.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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